Acknowledgments

We thank the US Embassy of Yaoundé, staff of Walter Reed Johns Hopkins Cameroon Program, T. Gerdes, goat owners, and the anonymous reviewers.

This research was financed by Johns Hopkins Bloomberg School of Public Health Center for a Livable Future (N.D.W.), the V. Kann Rasmussen Foundation (D.S.B.), the National Institutes of Health Fogarty International Center (grant #5 K01 TW000003-05 to N.D.W.), and an NIH Research Training Grant (S.U.). The Cameroon Ministry for Scientific and Technical Research provided research permits.

B. Matthew LeBreton,* Sally Umlauf,† Cyrille F. Djoko,*‡ Peter Daszak,§ Donald S. Burke,¶ Paul Yemgai Kwenkam,# and Nathan D. Wolfe¶

*Walter Reed Johns Hopkins Cameroon Program, Yaoundé, Cameroon; †Tufts University School of Veterinary Medicine, North Grafton, Massachusetts, USA; **‡**University of Yaoundé, Yaoundé, Cameroon; §The Consortium for Conservation Medicine, Palisades, New York, USA; ¶Johns Hopkins University, Baltimore, Maryland, USA; and #Ministry of Livestock, Fisheries and Animal Industries, Yaoundé, Cameroon

References

- Fontenille D, Traore-Lamizana M, Diallo M, Thonnon J, Digoutte JP, Zeller HG. New vectors of Rift Valley fever in West Africa. Emerg Infect Dis. 1998;4:289–93.
- Zeller HG, Bessin R, Thiongane Y, Bapetel I, Teou K, Ala MG, et al. 1995 Rift Valley fever antibody prevalence in domestic ungulates in Cameroon and several West African countries (1989–1992) following the 1987 Mauritanian outbreak. Res Virol. 1995;146:81–5.
- Ringot D, Durand J-P, Tolou H, Boutin J-P Davoust B. Rift Valley fever in Chad. Emerg Infect Dis. 2004;10:945–7.
- Nakounné E, Selekon B, Morvan J. Veille microbiologique : les fièvres hémorragiques virales en République Centrafricaine ; données sérologiques actualisées chez l'homme. Bull Soc Pathol Exot. 2000;93:340–7.

- Woods CW, Karpati AM, Grein T, McCarthy N, Gaturuku P, Muchiri E, et al. An outbreak of Rift Valley fever in northeastern Kenya, 1997–98. Emerg Infect Dis. 2002;8:138–44.
- Balkhy HH, Memish ZA. Rift Valley fever: an uninvited zoonosis in the Arabian peninsula. Int J Antimicrob Agents. 2003;21: 153–7.
- Paweska JT, Smith SJ, Wright IM, Williams R, Cohen AS, Van Dijk AA. Indirect enzyme-linked immunosorbent assay for the detection of antibody against Rift Valley fever virus in domestic and wild ruminant sera. Onderstepoort J Vet Res. 2003;70:49–64.
- Swanepoel R, Struthers JK, Erasmus MJ, Shepherd SP, McGillivry GM, Erasmus BJ, et al. Comparison of techniques for demonstrating antibodies to Rift Valley fever virus. J Hyg (Lond). 1986;97:317–29.
- Swanepoel R, Struther JK, Erasmus MJ, Shepherd SP, McGillivray GM, Shepherd AJ, et al. Comparative pathogenicity and antigenic cross-reactivity of Rift Valley fever and other African phleboviruses in sheep. J Hyg (Lond). 1986;97:331–46.
- Germani Y, Cunin P, Tedjouka E, Ncharre CB, Morvan J, Martin P. Enterohaemorrhagic *Escherichia coli* in Ngoila (Cameroon) during an outbreak of bloody diarrhoea. Lancet. 1998;352:625–6.

Address for correspondence: B. Matthew LeBreton, Walter Reed Johns Hopkins Cameroon Program; CRESAR; BP 7039; Yaoundé; Cameroon; fax: 237-221-3382; email: mlebreton@hopkinscameroon.org

HIV-1 CRF07_BC Infections, Injecting Drug Users, Taiwan

To the Editor: To date, Taiwan's human immunodeficiency virus type 1 (HIV-1) epidemic has primarily spread via sexual contact. The sub-type B and circulating recombinant form (CRF) 01_AE account for >95% of all infections (*1*). However, since

2003 Taiwan has experienced a major outbreak of CRF07_BC among injecting drug users (IDUs).

The first wave of HIV-1 infections in Taiwan can be traced to the early 1980s, when a group of hemophilia patients received imported HIV-1contaminated antihemophilia medications. By the time these medications had been replaced by heat-treated factor VIII concentrates, at least 53 patients had contracted HIV-1 infections (2). According to Taiwan's Center for Disease Control (CDC), HIV infections have been diagnosed in 9,229 persons (including 523 foreigners) as of July 31, 2005 (3). The number of persons living with HIV-1/AIDS has increased rapidly in the past few years, with a 77% increase in 2004, compared to 11% in 2003 (online Table, available at http:// www.cdc.gov/ncidod/EID/vol12no04 /05-0762.htm#table). According to the results of a risk factor analysis of people living with HIV-1/AIDS reported to the Taiwan CDC, the proportion of IDUs increased from 1.7% (13/773) in 2002 to 8.1% (70/861) in 2003 to 30.3% (462/1,521) in 2004 (online Table). The Taiwan CDC received reports of 1,241 IDUs diagnosed with HIV-1 infections from January 1 to July 31, 2005; these account for >75% of all reported HIV-1 infections in 2005(3). The evidence points to an explosive epidemic of HIV-1 infections among IDUs in Taiwan since 2003, with no indication of a slowdown.

Taiwan has $\approx 60,000$ IDUs (1). According to the Republic of China Ministry of Justice, the number of incarcerated drug offenders increased from 5,988 in 2003 to 9,303 in 2004; the rate of HIV-1 seropositive inmates increased from 13.3/100,000 in 2002 to 56.8/100,000 in 2004 (Y-M. Wu, Ministry of Justice, pers. comm.). Since all inmates are routinely tested for HIV-1 in detention centers, and all infected inmates are separated from HIV-1–seronegative inmates, the

LETTERS

potential of HIV-1 transmission in prisons is remote. We therefore suggest that the Taiwanese IDU population and its HIV-1 seropositive rate have both increased rapidly in the past few years.

To identify the primary HIV-1 strains in the current epidemic, we collected blood specimens from HIV-1-infected inmates in 3 detention centers (1 each located in the northern, central and southern regions of Taiwan). HIV-1 subtypes were determined by polymerase chain reaction, DNA sequencing, and phylogenetic analyses of pol or env genes. Our results indicate that 145 (96%) of 151 IDUs were infected with CRF07_BC and 6 (4%) were infected with subtype B; 97% of the CRF07 BC cases were diagnosed in 2003 or 2004. According to our phylogenetic analysis of the env gene, the Taiwanese CRF07_BC strains clustered with CRF07_BC strains drawn from IDUs in China (Figure).

CRF07_BC is a recombinant of the B' and C subtypes. Several studies have suggested that CRF07_BC originated in China's Yunnan Province, with subtype B' from Thailand mixing with subtype C from India before moving northwestward to Xinjiang Province along a major Chinese heroin trafficking route (4-6). To our knowledge, this is the first report of a large group of IDUs in northeastern Asia having CRF07 BC infections. It may have followed another drug trafficking route from Yunnan Province to southeast China, moving through Guangxi Province and Hong Kong to Taiwan (7–9). In a bid to combat skyrocketing HIV/AIDS infection rates among IDUs, the Taiwan CDC has proposed a 5-year harm reduction program to the Republic of China Executive Yuan.

Acknowledgments

We thank Yi-Ru Chang for her technical assistance, the Sequencing Core Facility of National Yang-Ming

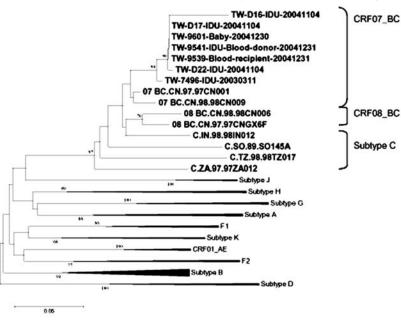


Figure. Phylogenetic analyses of 7 HIV-1 isolates identified in Taiwan. TW-7496, TW-D16, TW-D17, and TW-D22 were collected from detention center inmates; TW-9541 and TW-9539 were collected from a blood donor and 1 of his donation recipients. This neighborjoining tree was created from 100 bootstrap samples of aligned *env* sequences corresponding to the 7077–7340 nucleotide residues of HIV-1-HXB2 from different isolates. Bootstrap values are shown on branch nodes. Reference isolates from the GenBank HIV database are indicated by subtype. University's Genome Research Center for its technical assistance, and professor Kathleen Ahrens of National Taiwan University for her help in preparing this letter.

This research is partially supported by funds from the AIDS Molecular Virology Laboratory Program of the Taiwan CDC (CDC-94-RM-102, DOH93-DC-1043) and the ROC National Science Council's National Research Program for Genomic Medicine (95-0324-19-F-01-00-00-00-35).

Yi-Ming Arthur Chen,* Yu-Ching Lan,* Shu-Fen Lai,* Jyh-Yuan Yang,† Su-Fen Tsai,† and Steve Hsu-Sung Kuo†

*National Yang-Ming University, Taipei, Taiwan, Republic of China; and †Center for Disease Control, Taipei, Taiwan, Republic of China

References

- Chen YM, Huang KL, Jen I, Chen SC, Liu YC, Chuang YC, et al. Temporal trends and molecular epidemiology of HIV-1 infection in Taiwan from 1988 to 1998. J Acquir Immune Defic Syndr. 2001;26:274–82.
- Shen MC, Liu FY, Kuo YP, Hsie RF. A longitudinal study of immunological status in Chinese hemophiliacs: importance of the heat viral inactivation of factor concentrates. I. Immunological associations with the consumption of factor concentrates. Hemophilia. 1995;1:243–8.
- 3. Republic of China Center for Disease Control, Department of Health. Reported cases of HIV/AIDS by year in Taiwan 1984–2004. 2005. [cited 2005 Jul 31]. Available from http://www.cdc.gov.tw/en/ index/asp
- 4. Su L, Graf M, Zhang Y, von Briesen H, Xing H, Köstler J, et al. Characterization of a virtually full-length human immunodeficiency virus type 1 genome of a prevalent intersubtype (C/B') recombinant strain in China. J Virol. 2000;74:11367–76.
- Yang R, Xia X, Kusagawa S, Zhang C, Ben K, Takebe Y. On-going generation of multiple forms of HIV-1 intersubypte recombinants in the Yunnan province of China. AIDS. 2002;16:1401–7.
- Zhang L, Chen Z, Cao Y, Yu J, Li G, Yu W, et al. Molecular characterization of human immunodeficiency virus type 1 and hepatitis C virus in paid blood donors and infecting drug users in China. J Virol. 2004;78:13591–9.

- Piyasirisilp S, McCutchan FE, Carr JK, Anders-Buell E, Liu W, Chen J, et al. A recent outbreak of human immunodeficiency virus type 1 infection in southern China was initiated by two highly homogeneous geographically separated strains, circulating recombinant form AE and a novel BC recombinant. J Virol. 2000;74:11286–95.
- Cohen J. Asia and Africa: on different trajectories? Science. 2004;304:1932–8.
- Lim WL, Xing H, Wong KH, Wong MC, Shao YM, Ng MH, et al. The lack of an epidemiological link between HIV type 1 infections in Hong Kong and Mainland China. AIDS Res Hum Retroviruses. 2004;20:259–62.

Address for correspondence: Yi-Ming Arthur Chen, AIDS Prevention and Research Center, National Yang-Ming University, 155 Li-Nong St, Sec. 2, Taipei, Taiwan 112; fax: 886-2-2827-0576; email: arthur@ym.edu.tw

Chlamydialike Organisms and Atherosclerosis

To the Editor: Chlamydophila pneumoniae causes pneumonia, but its role in the pathogenesis of atherosclerosis is controversial (1-4). The role of C. pneumoniae in atherosclerosis is supported by seroepidemiologic studies and detection in atherosclerotic lesions by polymerase chain reaction (PCR), immunohistologic analysis, culture, and electron microscopy (2,3). However, these results were not confirmed by other serologic or PCR-based studies (4). Meijer et al. evaluated abdominal aortic aneurysm biopsy specimens and detected C. pneumoniae membrane antigen more frequently than lipopolysaccharide antigens but did not detect heat shock protein 60 (1). In addition, they could not amplify or detect specific C. pneumoniae DNA by PCR and fluorescence in situ hybridization (1). They

hypothesized that this discrepancy may result from a chlamydialike organism present in aortic samples that has surface antigens similar to those of *C. pneumoniae*.

Parachlamydia acanthamoebae and Neochlamydia hartmanellae are chlamydialike organisms that share ≈86% 16S rRNA sequence similarity with C. pneumoniae (5). Like C. pneumoniae, they have elementary and reticulate bodies visible by electron microscopy (6). Neochlamydiarelated DNA (GenBank accession no. AF097191) has been amplified from 5 different arterial samples, including 1 aortic aneurysm (7), and a relationship (p = 0.009) between cerebral hemorrhage and serologic evidence of Parachlamydia infection has been reported (8). Therefore, we investigated the role of Parachlamydia in pathogenesis of atherosclerosis by using a molecular approach.

We analyzed 78 surgical samples from 27 patients undergoing aortic or carotid surgery for atherosclerotic disease at Hôpital Nord in Marseille from June 1, 2003, to December 31, 2003. The study was approved by the local ethics committee, and written informed consent was obtained from all participants. Demographic and clinical data were prospectively recorded.

DNA was extracted from aortic or carotid samples with atherosclerotic lesions by using the QIAamp DNA tissue kit (Qiagen, Courtaboeuf, France), according to the manufacturer's instructions. A nested PCR was performed by using external primers 16SIGF (5'-CGGCGTGGATGAG-GCAT-3') and 16SIGR (5'-TCAGTC-CCAGTGTTGGC-3') (9) and internal primers CHL16SFOR2 (5'-CGTG-GATGAGGCATGCAAGTCGA-3') and CHL16SREV2 (5'-CAATCTCT-CAATCCGCCTAGACGTCTTAG-3') (7). PCR included negative controls from the DNA extraction step. DNA extractions and PCR amplifications were conducted in a laboratory in which parachlamydial DNA had not been extracted or amplified. PCR products were purified by using the QIAquick PCR purification kit (Qiagen) and sequenced by using the d-rhodamine terminator cycle sequencing reaction kit (Perkin-Elmer Biosystems, Warrington, UK) and a 3100 ABI Prism automated sequencer (Applied Biosystems, Courtaboeuf, France).

Sequences were analyzed with BLAST (http://www.ncbi.nlm.nih. gov/BLAST/) using gap existence and extension penalties of 5 and 2, respectively. Results were considered positive only when the sequence of the amplified product exhibited a best BLAST hit with a chlamydialike organism. Statistical analyses were performed with STATA software (Stata Corporation, College Station, TX, USA).

A positive PCR result was obtained with samples from 5(18.5%)of 27 patients (Table). Three sequences had a best BLAST hit with the sequence of Parachlamydia sp. UV7 (GenBank accession no. AJ715410), with a sequence similarity ranging from 99% to 100%. The other 2 sequences had 98% sequence similarity with Neochlamydia-related symbiont TUME-1 (GenBank accession no. AF098330). PCR positivity was not associated with age, sex, or location of the atherosclerotic lesion.

All patients with positive PCR results were ≥ 68 years of age. Patients without cardiovascular risk factors were more likely than those with ≥ 1 risk factor to have positive PCR results (p = 0.023). Despite the small number of patients in this study, this association was also confirmed in a multivariate logistic regression model adjusted for sex and previous cardiovascular disease (odds ratio 0.035, 95% confidence interval 0.001–0.94).

These findings suggest that *Parachlamydia* and *Neochlamydia* are associated with atherosclerosis. In addition, these obligate intracellular